

PREVALENCE OF PERIODONTAL PATHOGENS AMONG ITALIAN PATIENTS WITH CHRONIC PERIODONTITIS: A RETROSPECTIVE STUDY ON 2992 PATIENTS

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SUMMARY

Purpose. The aim of the present study was to evaluate the prevalence of some periodontal pathogens in Italian adults with chronic periodontitis.

Materials and methods. The sample consisted of 2992 patients with a clinical diagnosis of chronic periodontitis, based on the criteria of the American Academy of Periodontology, sampled in the period 2013-2016: 2108 patients were from Northern, 690 from Central and 194 from Southern Italy. *Porphyromonas gingivalis*, *Treponema denticola*, and *Tannerella forsythia* were investigated in all patients of the present study, while *Campylobacter rectus*, *Fusobacterium nucleatum* and *Aggregatibacter actinomycetemcomitans* only in 2514 (84%) patients.

Subgingival plaque samples of the four sites of greatest probing depth in each patient were used to obtain subgingival microbiota and then processed by quantitative polymerase chain reaction.

Results. Periodontal pathogens had the following presence respect to all amount of patients: *Aggregatibacter actinomycetemcomitans* 16.1%, *Campylobacter rectus* 73.4%, *Fusobacterium nucleatum* 93.8%, *Porphyromonas gingivalis* 65.5%, *Treponema denticola* 66.4%, and *Tannerella forsythia* 72.7%. There are no significant statistical differences among geographic areas both for the total bacterial and the single species except for *T. Denticola* and *C. Rectus*, which prevalence was significantly higher in Southern Italy (P value <.05). The other investigated species were equally distributed among different regions. *A. actinomycetemcomitans* was the rarer species detected in this study, while *F. nucleatum* was the commonest. No differences among areas were observed as regard of the mean bacterial load except for *F. Nucleatum* whose prevalence in Northern Italy was lower then both in Central and Southern Italy (P value <.05).

Conclusions. The results of our study didn't show different geographic distribution of periodontal pathogens among Italian population of the three areas investigated. The homogeneity of the results could be related to genetic and environmental factors.

Key words: periodontitis, pathogens, bacteria, population, oral disease.

Introduction

Periodontal disease (PD) is one of the most prevalent chronic diseases in the world and in the Italian population. Periodontal disease (PD) is characterized as a chronic inflammatory condition affecting the periodontal tissues and den-

tal plaque seems to play an essential role in the pathogenesis of this condition (1, 2).

PD is clearly caused by periodontal pathogens, in particularly some gram-negative anaerobic bacteria, such as *Aggregatibacter actinomycetemcomitans* (AA) and red complex bacteria (*Porphyromonas gingivalis*, PG -, *Tannerella forsythia*, TF and *Treponema denticola*, TD)

(3, 4). These gram-negative anaerobic bacteria have been demonstrated potential virulence factors, inducing host inflammatory mediators, leading to destruction of collagen of connective tissue due to specific enzymes, and alveolar bone resorption (5, 6).

The prevalence of periodontal pathogens is estimated very different in relation to geographic location and different populations (7, 8). Differences in prevalence and distribution of periodontal pathogens may play a major role in placing some population at a greater risk of infection and PD than others, so it is very important to know the epidemiology of PD for prevention and treatment of this disease (9, 10).

Presence and distribution of periodontal pathogens were investigated in Italian population (11-14). In these investigations, *Fusobacterium nucleatum* (FN) resulted the most frequently detected (95%) while TF showed the highest load. AA was the less represented bacterium for load and presence.

The aim of this investigation is to perform an epidemiological study about the presence and distribution of the most common bacteria in Italian adults with chronic PD.

Materials and methods

Patients

This epidemiological study was performed on patients of different private practices of North, Central and South of Italy between January 2013 and December 2016. The three areas included Italian regions according to Italian Institute of Statistics (ISTAT) (www.istat.it/it/archivio/regioni). The sample comprised 2992 patients: 2108 from North (70.0%), 690 from Central (23%) and 194 of South (7%) of Italy. *Porphyromonas gingivalis*, *Treponema denticola*, and *Tannerella forsythia* were investigated in all patients of the present study, while *Campylobacter rectus*, *Fusobacterium nucleatum* and *Aggregatibacter actino-*

mycetemcomitans only in 2514 (84%) of patients. All patients were diagnosed of PD according to AAP (American Academy of Periodontology) criteria, and selected on the basis of the following inclusion criteria: age >18 yrs., probing depth of 3 mm or more. These criteria state that the patient must have at least one site with a probing depth and clinical attachment loss ≥ 4 mm. Subgingival plaque samples of the four sites of greatest probing depth in each patient were used to obtain subgingival samples.

Microbiological evaluation

Samples for microbiological analysis were collected from the four sites of greatest probing depth in each quadrant, with sterilized n. 60 paper tips inserted to the depth of the pocket, left in place for 20 seconds, transferred to a sterile tube and sent for subsequent DNA extraction and polymerase chain reaction (PCR) analysis. Table 1 reported probe and primer sequences used for the amplification.

DNA extraction

After collection, paper probes were processed for bacterial DNA extraction, by using the GenElute™ Bacterial Genomic DNA Kit (Sigma-Aldrich, St., St. Louis, MO, USA) and following the manufacturing procedures. Briefly, to isolate DNA, samples were incubated with lysozyme in a specific lysis buffer and, subsequently with proteinase K. Later, extracted DNA was purified by spin-column method.

Real-Time Polymerase Chain Reaction

Real-Time Polymerase Chain Reaction using the absolute quantification with the standard curve method performed bacterial quantification.

Table 1 - Probe and primer sequences use for periodontal bacteria amplification.

Periodontal bacteria	Primer sequence 5' -> 3'	Probe sequence
Aa	f-ACCTTACCTACTCTTGACATCCGAA r-ATGCAGCACCTGTCTCAAAGC	AAGAACTCAGAGATGGGTTTGTGCCTAGG
Pg	f-CGCGTGAAGGAAGACAGTCC r-CGATGCTTATTCTTACGGTACATTCA	TACGGGAATAACGGGCGATACGAGTATTG
Tf	f-CAGCGATGGTAGCAATACCTGTC r- TTCGCCGGGTTATCCCTC	TGAGTAACGCGTATGTAACCTGCCCCG
Td	f-AGCTACGGCTCCGCTTCAG r-GATACCCATCGTTGCCTTGGT	AGCTAATGGGACGCGGGCCCCAT
Fn	f-AGGGTGATCGGCCACAAG r-CACAGAATTGCTGGATCAGACTCT	ACACGGCCCTTACTCCTACGGGAGG
Cr	f-TGACGCTAATGCGTGAAAGC r-CTCGACTAGCGAAGCAACAAC TAG	TACCCTGGTAGTCCACGCCCTAAACGA
TBL	f- TGGAGCATGTGGTTTAATTCGA r-TGCGGGACTTAACCCAACA	CACGAGCTGACGACARCCATGCA

Aa = *Aggregatibacter actinomycetemcomitans*; Pg = *Porphyromonas gingivalis*; Tf = *Tannerella forsythia*; Td = *Treponema denticola*; Fn = *Fusobacterium nucleatum*; Cr = *Campylobacter rectus*; TBL = Total bacterial load.

Primers and probes oligonucleotides for *P. gingivalis* and *T. forsythia* were designed basing on 16S rRNA gene sequences of the Human Oral Microbiome Database (HOMD 16S rRNA Ref-Seq Version 10.1).

For the quantitative analysis, plasmids (Eurofin MWG Operon, Ebersberg, Germany) containing the specific DNA target sequence were employed as standard.

All reactions were performed in duplex, in 20ul final volumes; with 2X TaqMan Universal PCR master mix (Applied Biosystems, Foster City, CA, USA) and 50nM concentration of each primers and 200nM of the probes. Amplifications were carried out by using the ABI PRISM 7500 (Applied Biosystems, Foster City, CA, USA).

Statistical analysis

Chi-square test was performed to compare the prevalence of each bacterial species in the patient groups. SPSS program was used to perform statistical tests. A 5% level of significance and 95% confidence interval were used for all tests.

Results

The observed load of each investigated bacterial species in the periodontal pockets of patients was reported in Table 2. *Porphyromonas gingivalis*, *Treponema denticola*, and *Tannerella forsythia* were investigated in all patients of the present study, while *Campylobacter rectus*, *Fusobacterium nucleatum* and *Aggregatibacter actinomycetemcomitans* only in 2514 (84%) patients. Periodontal pathogens have the following presence respect to all amount of patients: *Aggregatibacter actinomycetemcomitans* 16.1%, *Campylobacter rectus* 73.4%, *Fusobacterium nucleatum* 93.8%, *Porphyromonas gingivalis* 65.5%, *Treponema denticola* 66.4%, and *Tannerella forsythia* 72.7%. There are no significant differences in presence among the different geographic areas both for presence and distribution except for *T. Denticola* and *C. Rectus* which prevalence was significantly higher in Southern Italy. The other investigated species were equally distributed among different regions.

A. actinomycetemcomitans was the rarer species

Table 2 - Mean amounts of specific bacterial species detected in periodontal pockets of periodontitis patients.

Descriptive Statistics					
	N	Minimum	Maximum	Mean	Std. Deviation
AA	2514	,00	606731,00	2754,3500	21188,76
CBT	2992	65,00	4,2E+08	2897597	1,2E+07
CR	2514	,00	3909041	48174,84	181437,8
FN	2514	,00	9771033	243519,1	627376,0
PG	2992	,00	8976221	137964,1	506212,6
TD	2992	,00	2744233	74998,36	220887,8
TF	2992	,00	7735814	37262,02	206079,6
Valid N (listwise)	2514				

detected in this study, while *F. nucleatum* was the commonest. No differences among areas where observed as regard of the mean bacterial load except for *F. Nucleatum* whose prevalence in Northern Italy was lower than both Central and Southern Italy (Table 3, P value < 0.05).

Discussion

The present epidemiological study established the presence and distribution of PD bacteria in a large sample of Italian population, and investi-

Table 3 - Relative amounts of specific bacterial species detected in periodontal pockets of periodontitis patients.

		Report						
AREA		AA	CBT	CR	FN	PG	TD	TF
CENTRO	Mean	1984,4339	3681511	41738,89	282819,3	134343,6	83188,39	39160,13
	N	567	690	567	567	690	690	690
	Std. Deviation	17563,81	2,1E+07	153341,3	818138,4	511871,0	240731,5	193038,0
NORD	Mean	2717,1001	2621102	47578,83	223502,3	136695,3	72171,46	36620,55
	N	1759	2108	1759	1759	2108	2108	2108
	Std. Deviation	22392,82	7611808	185808,9	563732,3	478543,1	218703,5	217046,6
SUD	Mean	5424,9096	3113832	73161,82	312275,5	164627,7	76585,85	37481,19
	N	188	194	188	188	194	194	194
	Std. Deviation	19407,50	4651181	214227,9	512489,4	731455,8	163549,3	103988,5
Total	Mean	2754,3500	2897597	48174,84	243519,1	137964,1	74998,36	37262,02
	N	2514	2992	2514	2514	2992	2992	2992
	Std. Deviation	21188,76	1,2E+07	181437,8	627376,0	506212,6	220887,8	206079,6

Aa = *Aggregatibacter actinomycetemcomitans*; Pg = *Porphyromonas gingivalis*; Tf = *Tannerella forsythia*; Td = *Treponema denticola*; Fn = *Fusobacterium nucleatum*; Cr = *Campylobacter rectus*.

gates the differences among geographical areas. *AA* was found more rarely in Italian population, while *F. nucleatum* was the most frequent and in higher amount. The PD pathogens are related with the progression and severity of the disease. *F. nucleatum* is estimated one of the most abundant gram-negative anaerobes in patients with PD. The presence of *F. Nucleatum* is the main cause of PD worsening. The deeper is the pocket, more abundant is the amount of *F. Nucleatum*.

Differences in prevalence of TD and CR were observed. In addition difference in relative amounts of FN was detected among the three Italian areas. *F. Campilobacter Rectus* and *Treponema denticola* presence was significantly higher in Southern Italy. These results could be related to different genetic and environmental factors in the three areas. The study of presence and distribution of periodontal pathogens can be useful to tailor the best therapeutic protocol in each patient leaving in different geographic area (6, 15-21). Infection can happen with high frequencies in bone regeneration (22-25) also after cancer resection (26-30). In some pediatric conditions can be useful to have a low bacterial loading especially in syndromic conditions (31-33).

In addition, further studies should be performed to establish the relationship between periodontal pathogens and peri-implantitis. Tooth replacement with implants is a well-known technique used worldwide in the last 40 years (34-71). Bacteria of periodontal disease may worsen the implants survival rate (52, 53, 55, 72-86), or influence the surgical success in bone regeneration (22-25) or affecting young patients (87-92). Previous epidemiological studies in Italian population didn't report differences in PD pathogens presence and distribution. It may be explained by the homogeneity of periodontal diagnostic criteria. PD is one of the most diffuse chronic diseases in Italy, both in moderate to severe degree, and this fact explain the great amount of PD pathogens found in our study. In fact, the deeper is the pocket of the microbiological sample, the most aggressive are the bacteria, leading

to tooth mobility and loss.

In conclusion, our study showed there are no significant differences in presence and distribution among the different geographic areas except for *T. Denticola* and *C. Rectus* which prevalence was significantly higher in Southern Italy. In addition, we found a similar bacterial load in patients living in different Italian geographic areas, with the only exception of *F. Nucleatum*.

References

1. Lauritano D, Martinelli M, Mucchi D, Palmieri A, Muzio LL, Carinci F. Bacterial load of periodontal pathogens among Italian patients with chronic periodontitis: A comparative study of three different areas. *Journal of Biological Regulators and Homeostatic Agents*. 2016;30(2):149-54.
2. Lauritano D, Scapoli L, Mucchi D, Cura F, Muzio LLO, Carinci F. Infectogenomics: Lack of association between vdr, il6, il10 polymorphisms and "red Complex" bacterial load in a group of Italian adults with chronic periodontal disease. *Journal of Biological Regulators and Homeostatic Agents*. 2016;30(2):155-60.
3. Checchi L, Gatto MR, Checchi V, Carinci F. Bacteria prevalence in a large Italian population sample: A clinical and microbiological study. *Journal of Biological Regulators and Homeostatic Agents*. 2016;30(2):199-208.
4. Meynardi F, Pasqualini ME, Rossi F, Dal Carlo L, Biancotti P, Carinci F. Correlation between dysfunctional occlusion and periodontal bacterial profile. *J Biol Regul Homeost Agents*. 2016;30(2 Suppl 1):115-21.
5. Lombardo L, Carinci F, Martini M, Gemmati D, Nardone M, Siciliani G. Quantitive evaluation of dentin sialoprotein (DSP) using microbeads - A potential early marker of root resorption. *ORAL and Implantology*. 2016;9(3):132-42.
6. Lauritano D, Cura F, Candotto V, Gaudio RM, Mucchi D, Carinci F. Evaluation of the Efficacy of Titanium Dioxide with Monovalent Silver Ions Covalently Linked (Tiab) as an Adjunct to Scaling and Root Planing in the Management of Chronic Periodontitis Using Pcr Analysis: A Microbiological Study. *J Biol Regul Homeost Agents*. 2015;29(3 Suppl 1):127-30.
7. Quirynen M, van der Mei HC, Bollen CM, Schotte A, Marechal M, Doornbusch GI, Naert I, Busscher HJ, van Steenberghe D. An in vivo study of the influence of the surface roughness of implants on the microbiology of supra- and subgingival plaque. *J Dent Res*. 1993; 72(9):1304-9.

8. Lauritano D, Cura F, Candotto V, Gaudio RM, Mucchi D, Carinci F. Periodontal Pockets as a Reservoir of *Helicobacter Pylori* Causing Relapse of Gastric Ulcer: A Review of the Literature. *J Biol Regul Homeost Agents*. 2015;29(3 Suppl 1):123-6.
9. Scapoli L, Girardi A, Palmieri A, Martinelli M, Cura F, Lauritano D, Pezzetti F, Carinci F. Interleukin-6 Gene Polymorphism Modulates the Risk of Periodontal Diseases. *J Biol Regul Homeost Agents*. 2015;29(3 Suppl 1):111-6.
10. Carinci F, Girardi A, Palmieri A, Martinelli M, Scapoli L, Avantaggiato A, Nardi GM, Lauritano D. LAB®-Test 1: Peri-Implantitis and bacteriological analysis. *European Journal of Inflammation*. 2012;10(1):91-93.
11. Carinci F, Girardi A, Palmieri A, Martinelli M, Scapoli L, Avantaggiato A, Nardi GM, Lauritano D. LAB®-test 2: Microflora and periodontal disease. *European Journal of Inflammation*. 2012;10(1):95-98.
12. Carinci F, Girardi A, Palmieri A, Martinelli M, Scapoli L, Avantaggiato A, Nardi GM, Lauritano D. Lab®-test 3: Genetic susceptibility in periodontal disease. *European Journal of Inflammation*. 2012;10(1):99-101.
13. Scapoli L, Girardi A, Palmieri A, Carinci F, Testori T, Zuffetti F, Monguzzi R, Lauritano D. IL6 and IL10 are genetic susceptibility factors of periodontal disease. *Dent Res J (Isfahan)*. 2012;9(Suppl 2):S197-201.
14. Scapoli L, Girardi A, Palmieri A, Testori T, Zuffetti F, Monguzzi R, Lauritano D, Carinci F. Microflora and periodontal disease. *Dent Res J (Isfahan)*. 2012;9(Suppl 2):S202-6.
15. Cura F, Palmieri A, Girardi A, Martinelli M, Scapoli L, Carinci F. Lab-Test((R)) 4: Dental caries and bacteriological analysis. *Dent Res J (Isfahan)*. 2012;9(Suppl 2):S139-41.
16. Roncati M, Lauritano D, Cura F, Carinci F. Evaluation of light-emitting diode (led-835 nm) application over human gingival fibroblast: An in vitro study. *Journal of Biological Regulators and Homeostatic Agents*. 2016;30(2):161-67.
17. Caccianiga G, Rey G, Paiusco A, Lauritano D, Cura F, Ormianer Z, Carinci F. Oxygen high level laser therapy is efficient in treatment of chronic periodontitis: A clinical and microbiological study using PCR analysis. *Journal of Biological Regulators and Homeostatic Agents*. 2016;30(2):87-97.
18. Lauritano D, Bignozzi CA, Pazzi D, Palmieri A, Gaudio RM, Di Muzio M, Carinci F. Evaluation of the efficacy of a new oral gel as an adjunct to home oral hygiene in the management of chronic periodontitis. A microbiological study using PCR analysis. *J Biol Regul Homeost Agents*. 2016;30(2 Suppl 1):123-8.
19. Carinci F, Palmieri A, Girardi A, Cura F, Lauritano D. Aquolab® ozone-therapy is an efficient adjuvant in the treatment of chronic periodontitis: A case-control study. *Journal of Orofacial Sciences*. 2015;7(1):27-32.
20. Lauritano D, Cura F, Gaudio RM, Pezzetti F, Andreasi Bassi M, Carinci F. Polymerase Chain Reaction to Evaluate the Efficacy of Silica Dioxide Colloidal Solutions in the Treatment of Chronic Periodontitis: A Case Control Study. *J Biol Regul Homeost Agents*. 2015;29(3 Suppl 1):131-5.
21. Lauritano D, Petrucci M, Nardi GM, Carinci F, Minervini G, Di Stasio D, Lucchese A. Single Application of a Dessicating Agent in the Treatment of Recurrent Aphthous Stomatitis. *J Biol Regul Homeost Agents*. 2015;29(3 Suppl 1):59-66.
22. Mangano C, Piattelli A, Tettamanti L, Mangano F, Mangano A, Borges F, Iezzi G, d'Avila S, Shibli JA. Engineered bone by autologous osteoblasts on polymeric scaffolds in maxillary sinus augmentation: histologic report. *The Journal of oral implantology*. 2010;36(6):491-96.
23. Ballini A, Mastrangelo F, Gastaldi G, Tettamanti L, Bukvic N, Cantore S, Cocco T, Saini R, Desiate A, Gherlone E, Scacco S. Osteogenic differentiation and gene expression of dental pulp stem cells under low-level laser irradiation: a good promise for tissue engineering. *Journal of biological regulators and homeostatic agents*. 2015;29(4):813-22.
24. Mangano FG, Tettamanti L, Sammons RL, Azzi L, Caprioglio A, MacChi A, Mangano C. Maxillary sinus augmentation with adult mesenchymal stem cells: A review of the current literature. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*. 2013;115(6):717-23.
25. Mastrangelo F, Quaresima R, Grilli A, Tettamanti L, Vinci R, Sammartino G, Tetè S, Gherlone E. A comparison of bovine bone and hydroxyapatite scaffolds during initial bone regeneration: An in vitro evaluation. *Implant Dentistry*. 2013;22(6):613-22.
26. Carinci F, Stabellini G, Calvitti M, Pelucchi S, Targa L, Farina A, Pezzetti F, Pastore A. CD44 as prognostic factor in oral and oropharyngeal squamous cell carcinoma. *J Craniofac Surg*. 2002;13(1):85-9.
27. Mariani G, Calastrini C, Carinci F, Marzola R, Calura G. Ultrastructural features of cyclosporine A-induced gingival hyperplasia. *Journal of Periodontology*. 1993;64(11):1092-97.
28. Francioso F, Carinci F, Tosi L, Scapoli L, Pezzetti F, Passerella E, Evangelisti R, Pastore A, Pelucchi S, Piattelli A, Rubini C, Fioroni M, Carinci P, Volinia S. Identification of differentially expressed genes in human salivary gland tumors by DNA microarrays. *Molecular Cancer Therapeutics*. 2002;1(7):533-38.
29. Bodo M, Lilli C, Bellucci C, Carinci P, Calvitti M, Pezzetti F, Stabellini G, Bellocchio S, Balducci C, Carinci F, Baroni T. Basic fibroblast growth factor autocrine loop controls human osteosarcoma phenotyping and differentiation. *Molecular Medicine*. 2002;8(7):393-404.
30. Carinci F, Lo Muzio L, Piattelli A, Rubini C, Chiesa F, Ionna F, Palmieri A, Maiorano E, Pastore A, Laino G, Favia G, Dolci M, Pezzetti F. Potential markers of tongue tumor progression selected by cDNA microar-

- ray. International Journal of Immunopathology and Pharmacology. 2005;18(3):513-24.
31. Carinci F, Avantaggiato A, Curioni C. Crouzon syndrome: Cephalometric analysis and evaluation of pathogenesis. Cleft Palate-Craniofacial Journal. 1994; 31(3):201-09.
32. Bodo M, Carinci F, Baroni T, Giammarioli M, Bellucci C, Bosi G, Pezzetti F, Becchetti E, Evangelisti R, Carinci P. Apert's syndrome: Differential in vitro production of matrix macromolecules and its regulation by interleukins. European Journal of Clinical Investigation. 1997;27(1):36-42.
33. Martinelli M, Scapoli L, Palmieri A, Pezzetti F, Baciliero U, Padula E, Carinci P, Morselli PG, Carinci F. Study of four genes belonging to the folate pathway: transcobalamin 2 is involved in the onset of non-syndromic cleft lip with or without cleft palate. Human mutation. 2006;27(3):294.
34. Rigo L, Viscioni A, Franco M, Lucchese A, Zollino I, Brunelli G, Carinci F. Overdentures on implants placed in bone augmented with fresh frozen bone. Minerva Stomatol. 2011;60(1-2):5-14.
35. Carinci F, Brunelli G, Franco M, Viscioni A, Rigo L, Guidi R, Strohmer L. A retrospective study on 287 implants installed in resorbed maxillae grafted with fresh frozen allogeneous bone. Clin Implant Dent Relat Res. 2010;12(2):91-8.
36. Viscioni A, Rigo L, Franco M, Brunelli G, Avantaggiato A, Sollazzo V, Carinci F. Reconstruction of severely atrophic jaws using homografts and simultaneous implant placement: a retrospective study. J Oral Implantol. 2010;36(2):131-9.
37. Franco M, Rigo L, Viscione A, De Santis B, Tropina E, Brunelli G, Guidi R, Avantaggiato A, Carinci F. CaPO4 blasted implants inserted into iliac crest homologue frozen grafts. The Journal of oral implantology. 2009; 35(4):176-80.
38. Viscioni A, Franco M, Rigo L, Guidi R, Brunelli G, Carinci F. Implants inserted into homografts bearing fixed restorations. Int J Prosthodont. 2009;22(2):148-54.
39. Franco M, Viscioni A, Rigo L, Guidi R, Zollino I, Avantaggiato A, Carinci F. Clinical outcome of narrow diameter implants inserted into allografts. J Appl Oral Sci. 2009;17(4):301-6.
40. Viscioni A, Franco M, Rigo L, Guidi R, Spinelli G, Carinci F. Retrospective study of standard-diameter implants inserted into allografts. J Oral Maxillofac Surg. 2009;67(2):387-93.
41. Carinci F, Brunelli G, Zollino H, Franco M, Viscioni A, Rigo L, Guidi R, Strohmer L. Mandibles grafted with fresh-frozen bone: An evaluation of implant outcome. Implant Dentistry. 2009;18(1):86-95.
42. Carinci F, Brunelli G, Zollino I, Franco M, Viscioni A, Rigo L, Guidi R, Strohmer L. Mandibles grafted with fresh-frozen bone: an evaluation of implant outcome. Implant Dent. 2009;18(1):86-95.
43. Franco M, Tropina E, De Santis B, Viscioni A, Rigo L, Guidi R, Carinci F. A 2-year follow-up study on standard length implants inserted into alveolar bone sites augmented with homografts. Stomatologija. 2008;10(4):127-32.
44. Lucchese A, Carinci F, Saggese V, Lauritano D. Immediate loading versus traditional approach in functional implantology. European Journal of Inflammation. 2012;10(1 SUPPL. 3):55-58.
45. Traini T, Danza M, Zollino I, Altavilla R, Lucchese A, Sollazzo V, Trapella G, Brunelli G, Carinci F. Histomorphometric evaluation of an immediately loaded implant retrieved from human mandible after 2 years. International Journal of Immunopathology and Pharmacology. 2011;24(31-36).
46. Scarano A, Murmura G, Carinci F, Lauritano D. Immediately loaded small-diameter dental implants: evaluation of retention, stability and comfort for the edentulous patient. European Journal of Inflammation. 2012;10(1 S2):19-23.
47. Degidi M, Piattelli A, Carinci F. Clinical outcome of narrow diameter implants: a retrospective study of 510 implants. J Periodontol. 2008;79(1):49-54.
48. Degidi M, Piattelli A, Iezzi G, Carinci F. Do longer implants improve clinical outcome in immediate loading? Int J Oral Maxillofac Surg. 2007;36(12):1172-6.
49. Degidi M, Piattelli A, Carinci F. Immediate loaded dental implants: comparison between fixtures inserted in postextractive and healed bone sites. J Craniofac Surg. 2007;18(4):965-71.
50. Degidi M, Piattelli A, Iezzi G, Carinci F. Retrospective study of 200 immediately loaded implants retaining 50 mandibular overdentures. Quintessence Int. 2007;38(4):281-8.
51. Degidi M, Piattelli A, Iezzi G, Carinci F. Immediately loaded short implants: analysis of a case series of 133 implants. Quintessence Int. 2007;38(3):193-201.
52. Degidi M, Piattelli A, Iezzi G, Carinci F. Wide-diameter implants: Analysis of clinical outcome of 304 fixtures. Journal of Periodontology. 2007;78(1): 52-58.
53. Degidi M, Piattelli A, Gehrke P, Felice P, Carinci F. Five-year outcome of 111 immediate nonfunctional single restorations. J Oral Implantol. 2006;32(6):277-85.
54. Degidi M, Piattelli A, Carinci F. Parallel screw cylinder implants: Comparative analysis between immediate loading and two-stage healing of 1005 dental implants with a 2-year follow up. Clinical Implant Dentistry and Related Research. 2006;8(3):151-60.
55. Degidi M, Piattelli A, Gehrke P, Carinci F. Clinical outcome of 802 immediately loaded 2-stage submerged implants with a new grit-blasted and acid-etched surface: 12-month follow-up. Int J Oral Maxillofac Implants. 2006;21(5):763-8.
56. Degidi M, Piattelli A, Felice P, Carinci F. Immediate functional loading of edentulous maxilla: a 5-year ret-

- respective study of 388 titanium implants. *J Periodontol.* 2005;76(6):1016-24.
57. Danza M, Paracchini L, Carinci F. Tridimensional finite element analysis to detect stress distribution in implants. *Dental Cadmos.* 2012;80(10):598-602.
 58. Danza M, Grecchi F, Zollino I, Casadio C, Carinci F. Spiral implants bearing full-arch rehabilitation: Analysis of clinical outcome. *Journal of Oral Implantology.* 2011;37(4):447-55.
 59. Danza M, Zollino I, Avantaggiato A, Lucchese A, Carinci F. Distance between implants has a potential impact of crestal bone resorption. *Saudi Dental Journal.* 2011;23(3):129-33.
 60. Carinci F, Danza M. Clinical outcome of implants inserted in piezo split alveolar ridges: A pilot study. In: *Perspectives on Clinical Dentistry.* 2011:29-30.
 61. Danza M, Zollino I, Guidi R, Carinci F. Computer planned implantology: Analysis of a case series. In: *Perspectives on Clinical Dentistry.* 2011:287-300.
 62. Danza M, Carinci F. Flapless surgery and immediately loaded implants: a retrospective comparison between implantation with and without computer-assisted planned surgical stent. *Stomatologija.* 2010;12(2):35-41.
 63. Danza M, Quaranta A, Carinci F, Paracchini L, Pompa G, Voza I. Biomechanical evaluation of dental implants in D1 and D4 bone by Finite Element Analysis. *Minerva stomatologica.* 2010;59(6):305-13.
 64. Danza M, Riccardo G, Carinci F. Bone platform switching: a retrospective study on the slope of reverse conical neck. *Quintessence Int.* 2010;41(1):35-40.
 65. Danza M, Fromovich O, Guidi R, Carinci F. The clinical outcomes of 234 spiral family implants. *J Contemp Dent Pract.* 2009;10(5):E049-56.
 66. Calvo-Guirado JL, Ortiz-Ruiz AJ, Lopez-Mari L, Delgado-Ruiz R, Mate-Sanchez J, Bravo Gonzalez LA. Immediate maxillary restoration of single-tooth implants using platform switching for crestal bone preservation: a 12-month study. *Int J Oral Maxillofac Implants.* 2009;24(2):275-81.
 67. Danza M, Guidi R, Carinci F. Comparison Between Implants Inserted Into Piezo Split and Unsplit Alveolar Crests. *Journal of Oral and Maxillofacial Surgery.* 2009;67(11):2460-65.
 68. Danza M, Scarano A, Zollino I, Carinci F. Evaluation of biological width around implants inserted in native alveolar crest bone. *Journal of Osseointegration.* 2009;1(2):73-76.
 69. Danza M, Zollino I, Guidi R, Carinci F. A new device for impression transfer for non-parallel endosseous implants. *Saudi Dental Journal.* 2009;21(2):79-81.
 70. Andreasi Bassi M, Lopez MA, Confalone L, Gaudio RM, Lombardo L, Lauritano D. Clinical outcome of a two-piece implant system with an internal hexagonal connection: a prospective study. *J Biol Regul Homeost Agents.* 2016;30(2 Suppl 1):7-12.
 71. Danza M, Guidi R, Carinci F. Spiral family implants inserted in postextraction bone sites. *Implant Dent.* 2009;18(3):270-8.
 72. Carinci F, Lauritano D, Cura F, Lopez MA, Bassi MA, Confalone L, Pezzetti F. Prevention of bacterial leakage at implant-Abutment connection level: An in vitro study of the efficacy of three different implant systems. *Journal of Biological Regulators and Homeostatic Agents.* 2016;30(2):69-73.
 73. El Haddad E, Gianni AB, Mancini GE, Cura F, Carinci F. Implant-abutment leaking of replace conical connection nobel biocare® implant system. An in vitro study of the microbiological penetration from external environment to implant-abutment space. *ORAL and Implantology.* 2016;9(2):76-82.
 74. Mancini GE, Gianni AB, Cura F, Ormanier Z, Carinci F. Efficacy of a new implant-abutment connection to minimize microbial contamination: An in vitro study. *ORAL and Implantology.* 2016;9(3):99-105.
 75. Roncati M, Lucchese A, Carinci F. Non-Surgical treatment of peri-Implantitis with the adjunctive use of an 810-nm diode laser. *Journal of Indian Society of Periodontology.* 2013;17(6):812-15.
 76. Scarano A, Tripodi D, Carinci F, Piccolomini R, D'Ercole S. Biofilm formation on titanium alloy and anatase-Bactercline® coated titanium healing screws: An in vivo human study. *Journal of Osseointegration.* 2013;5(1):8-12.
 77. Brunelli G, Carinci F, Zollino I, Candotto V, Scarano A, Lauritano D. Sem evaluation of 10 infected implants retrieved from man. *European Journal of Inflammation.* 2012;10(1 S2):7-12.
 78. Scarano A, Sinjari B, Di Orio D, Murmura G, Carinci F, Lauritano D. Surface analysis of failed oral titanium implants after irradiated with ErCr:ysgg 2780 laser. *European Journal of Inflammation.* 2012;10(1 S2):49-54.
 79. Brunelli G, Carinci F, Zollino I, Candotto V, Scarano A, Lauritano D. Peri-implantitis. A case report and literature review. *European Journal of Inflammation.* 2012;10(1 S2):1-5.
 80. Scarano A, Piattelli A, Polimeni A, Di Iorio D, Carinci F. Bacterial adhesion on commercially pure titanium and anatase-coated titanium healing screws: An in vivo human study. *Journal of Periodontology.* 2010;81(10):1466-71.
 81. Grecchi F, Zollino I, Candotto V, Gallo F, Rubino G, Giglio S, Bianco R, Carinci F. A case of mandible osteonecrosis after a severe periimplant infection. *Dent Res J (Isfahan).* 2012;9(Suppl 2):S233-6.
 82. Carinci F, Farina A, Zanetti U, Vinci R, Negrini S, Calura G, Laino G, Piattelli A. Alveolar ridge augmentation: a comparative longitudinal study between calvaria and iliac crest bone grafts. *J Oral Implantol.* 2005;31(1):39-45.
 83. Carinci F, Pezzetti F, Volinia S, Francioso F, Arcelli D, Marchesini J, Caramelli E, Piattelli A. Analysis of MG63 osteoblastic-cell response to a new nanoporous

- implant surface by means of a microarray technology. *Clinical Oral Implants Research*. 2004;15(2):180-86.
84. Oliveira DP, Palmieri A, Carinci F, Bolfarini C. Gene expression of human osteoblasts cells on chemically treated surfaces of Ti-6Al-4V-ELI. *Mater Sci Eng C Mater Biol Appl*. 2015;51(248-55).
85. Andreasi Bassi M, Lopez MA, Confalone L, Carinci F. Hydraulic sinus lift technique in future site development: clinical and histomorphometric analysis of human biopsies. *Implant Dent*. 2015;24(1):117-24.
86. El Haddad E, Lauritano D, Carinci F. Interradicular septum as guide for pilot drill in postextractive implantology: a technical note. *J Contemp Dent Pract*. 2015;16(1):81-4.
87. Spadari F, Venesia P, Azzi L, Veronesi G, Costantino D, Croveri F, Farronato D, Tagliabue A, Tettamanti L. Low basal salivary flow and Burning Mouth Syndrome: New evidence in this enigmatic pathology. *Journal of Oral Pathology and Medicine*. 2015;44(3):229-33.
88. Tettamanti L, Caprioglio A, Tecco S, Barelo G, Macchi A, Tagliabue A, Levrini L. Oral squamous cell carcinoma in the paediatric patient: A literature review. *European Journal of Paediatric Dentistry*. 2012;13(1):35-40.
89. Caprioglio A, Mariani L, Tettamanti L. A pilot study about emotional experiences by using CFSS-DS in young patients. *European journal of paediatric dentistry: official journal of European Academy of Paediatric Dentistry*. 2009;10(3):121-24.
90. Levrini L, Tettamanti L, Abbate GM, Caria MP, Caprioglio A. pH of tooth surface in healthy adolescents at rest and after a glucose rinse: Effect of 72 hours of plaque accumulation. *European Journal of Paediatric Dentistry*. 2012;13(4):293-96.
91. Brenna F, Tagliabue A, Levrini L, Tettamanti L, Quacci D, Bergamaschi M. Scanning electron microscopy evaluation of the link between a new dentinal desensitizer and dentine. *Minerva stomatologica*. 2003;52(9):
92. Levrini L, Tettamanti L, Macchi A, Tagliabue A, Caprioglio A. Invisalign teen for thumb-sucking management. A case report. *European Journal of Paediatric Dentistry*. 2012;13(2):155-58.

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